

EFFICACY OF CEILING MOUNTED UV-C SYSTEMS



UVANGEL®
PATHOGEN CONTROL TECHNOLOGY

The CDC states that ultraviolet irradiation of air is an effective means of "reducing the transmission of airborne bacterial and viral infections in hospitals."¹ Ultraviolet germicidal irradiation (UVGI) occurs when UV light at an effective wavelength of 254 nanometers, disrupts the nucleic acid in the DNA of a microorganism, preventing it from replicating.

The development of active UVGI air treatment systems that assume the footprint of a standard 2' x 4' ceiling panel or light fixture was developed in recent years. Similar to upper room air treatment and active air duct treatment, these systems can be safely used in occupied spaces 24/7/365 where the pathogens are generated and freely circulated. Below are a few studies that show the effectiveness of this UV-C technology in healthcare.

PEER-REVIEW HOSPITAL

After installing UV-C ceiling mounted systems, airborne bacteria in patient rooms were reduced an average of **42%** in a hospital in Kentucky.² Common HAIs and catheter-associated urinary tract infections were reduced significantly as were overall infections by **60%**. There were no reported changes to the amount or type of cleaning done, infection control protocols, or reporting procedures. Other infections traditionally considered contact transmissible (central line-associated bloodstream infection and methicillin-resistant *Staphylococcus aureus*), also declined noticeably.

C. Diff reduced 88%
MRSA reduced 54%
VREs reduced 14%

CAUTIs reduced 55%
CLABSIs reduced 44%
Overall infections reduced 60%

Conclusions: Continuous shielded UV-C reduced airborne bacteria and may also lower the number of HAIs, including those caused by contact pathogens. Reduced infections result in lessened morbidity and lower costs.

PEER-REVIEW LTCF

Over the course of six months, data was collected and analyzed in a study that was conducted at a long-term care hospital in TN.³ The overall infection rate was significantly lower in rooms with UV-C units than in those without. The bacteria air sampling in the patient rooms were reduced by **51%** and the total reduction in infections dropped by **28%**. An anecdotal note to this study, staff reported that allergy symptoms were reduced, and absenteeism was lowest in the wing where the UV-C systems were installed.

Airborne bacteria reduced 51% Infection rate reduced 28%

Conclusion: Findings suggest that continuous exposure to UV-C treated air reduces HAIs. Shielded UV-C units in patient rooms may be an effective non-staff intervention dependent method for reducing HAIs.

PEER-REVIEW PHARMACY STUDY

Viable air particles pose a risk in areas where sterile preparations are compounded.⁴ Mean airborne fungal and bacterial colony forming units were obtained pre-installation and again in 6 months. A statistically significant decrease of **78%** and **62%** was observed for fungal and bacterial particles, respectively.

After installing the UV-C systems in the anteroom, dispensing/receiving and processing areas, bacteria and fungi was decreased in the anteroom by **86%** and **90%** respectively. The UV-C systems reduced the contaminated air flow, so the levels of bacteria and fungi were decreased by **92%** and **100%** in the compounding IV room where no units were installed.

| Anteroom Results | Pre-CFUs | Post-CFUs | % Decrease |
|-----------------------|----------|-----------|------------|
| Fungi Air Sampling | 1.8 | 0.18 | 90% |
| Bacteria Air Sampling | 35.3 | 4.85 | 86% |
| Compounding Results | | | |
| Fungi Air Sampling | 3.25 | 0.0 | 100% |
| Bacteria Air Sampling | 1.5 | 0.125 | 92% |

Conclusions: This study demonstrates how using shielded UV-C technology can decrease the spread of airborne pathogens throughout a compounding pharmacy.

PEER REVIEW AIR AND SURFACE

Field trials were set up at three hospitals (Texas, Nevada, and Massachusetts) where we tested air and surface for bacteria, installed continuous UV-C products at the room level, and then tested air and surface again.⁵ In all cases, airborne bacteria was reduced between **79%** and **91%** over pre-installation values. Most surfaces also showed reductions in bacteria from **48%** to **69%**, although we report one incident of an increase of 288%.

Conclusions: The data indicate that using active, shielded UV-C air technology at the room level reduces the bioburden in the air and on surfaces, including in occupied spaces.

UV vs. CORONAVIRUSES

There is currently great interest in emerging pathogens like coronaviruses. Approximately 100 sequences of the SARS-CoV-2 genome have been published and these suggest there are two types, Type I and Type II, of which the latter came from the Huanan market in China while the Type I strain came from an unknown location (Zhang 2020).

The effectiveness of UV on Coronaviruses was started by Hirano back in 1978. The table below summarizes the results of studies that have been performed on Coronaviruses under ultraviolet light exposure, with the specific species indicated in each case. The D90 value indicates the ultraviolet dose for 90% inactivation. Although there is a wide range of variation in the D90 values, this is typical of laboratory studies on ultraviolet susceptibility. The range of D90 values for coronaviruses is 7-241 J/m², the average which is 67 J/m², should adequately represent the ultraviolet susceptibility of the SARS-CoV-2 (COVID-19) virus.

Table 1: Summary of Ultraviolet Studies on Coronaviruses

| Microbe | D ₉₀ Dose J/m ² | UV k m ² /J | Base Pairs kb | Source |
|-----------------------------|---------------------------------------|------------------------|---------------|----------------------------|
| Coronavirus | 7 | 0.35120 | 30741 | Walker 2007 ^a |
| Berne virus (Coronaviridae) | 7 | 0.32100 | 28480 | Weiss 1986 |
| Murine Coronavirus (MHV) | 15 | 0.15351 | 31335 | Hirano 1978 |
| Canine Coronavirus (CCV) | 29 | 0.08079 | 29278 | Saknimit 1988 ^b |
| Murine Coronavirus (MHV) | 29 | 0.08079 | 31335 | Saknimit 1988 ^b |
| SARS Coronavirus CoV-P9 | 40 | 0.05750 | 29829 | Duan 2003 ^c |
| Murine Coronavirus (MHV) | 103 | 0.02240 | 31335 | Liu 2003 |
| SARS Coronavirus (Hanoi) | 134 | 0.01720 | 29751 | Kariwa 2004 ^d |
| SARS Coronavirus (Urbani) | 241 | 0.00955 | 29751 | Darnell 2004 |
| Average | 67 | 0.03433 | | |

^a (Jingwen 2020)

^b (estimated)

^c (mean estimate)

^d (at 3 logs)

UV ANGEL PERFORMANCE/VALIDATION STUDIES

UV Angel has conducted two separate laboratory tests by an independent third party against surrogate pathogens including *Escherichia coli* (gram negative), *Staphylococcus aureus* (gram positive), *Cladosporium cladosporioides* (fungus spore formers) and MS2 Bacteriophage (MS2) (virus surrogate).⁶ The UV Angel Air showed elimination rates from **90%**. Laboratory tests and mathematical modeling show elimination rates approaching 100% against more than 80 serious disease-causing pathogens.

UV mathematical modeling and D90 rates have been established for 80 pathogens, known or suspected airborne component in their transmission cycle, including bacteria, viruses, and fungi. Many pathogens, if they are drawn into the UVGI chamber, are neutralized in a single pass. Perhaps more significantly, for some of the most virulent pathogens, including MRSA, VRE, and *C. difficile*, the removal rate (reflecting both filtration and UV disinfection) was 100 percent modeled for those pathogens that pass through the chamber.

Table 4: Combined UV + Filter Removal Rates

| Microbe | Type | Size μm | Filter % | UV Rate % | Total % |
|-----------------------------|----------|--------------------|----------|-----------|---------|
| Acinetobacter | Bacteria | 1.225 | 21 | 100 | 100.00 |
| Adenovirus | Virus | 0.079 | 9 | 100 | 100.00 |
| Aeromonas | Bacteria | 2.098 | 35 | 100 | 100.00 |
| Aspergillus | Fungi | 3.354 | 45 | 93 | 96.30 |
| Bacillus anthracis | Bacteria | 1.118 | 19 | 61 | 68.20 |
| Bacteroides fragilis | Bacteria | 3.162 | 44 | 100 | 100.00 |
| Blastomyces dermatitidis | Fungi | 12.649 | 50 | 99 | 99.65 |
| Bordetella pertussis | Bacteria | 0.245 | 4 | 100 | 100.00 |
| Burkholderia cenocepacia | Bacteria | 0.707 | 11 | 100 | 100.00 |
| Burkholderia mallei | Bacteria | 0.674 | 10 | 100 | 100.00 |
| Burkholderia pseudomallei | Bacteria | 0.494 | 7 | 100 | 100.00 |
| Candida albicans | Fungi | 4.899 | 49 | 79 | 89.19 |
| Candia auris | Fungi | 4.899 | 49 | 75 | 87.31 |
| Chlamydia pneumoniae | Bacteria | 0.548 | 8 | 100 | 100.00 |
| Chlamydia psittaci | Bacteria | 0.283 | 4 | 100 | 100.00 |
| Cladosporium | Fungi | 8.062 | 50 | 98 | 98.75 |
| Clostridium botulinum | Bacteria | 1.975 | 33 | 100 | 100.00 |
| Clostridium difficile | Bacteria | 2 | 34 | 100 | 100.00 |
| Clostridium perfringens | Bacteria | 5 | 49 | 100 | 100.00 |
| Coronavirus (Wuhan) | Virus | 0.11 | 6 | 100 | 100.00 |
| Corynebacterium diphtheriae | Bacteria | 0.698 | 10 | 100 | 100.00 |
| Coxsackievirus | Virus | 0.027 | 19 | 100 | 100.00 |
| Cryptococcus neoformans | Fungi | 4.899 | 49 | 99 | 99.67 |
| Curvularia lunata | Fungi | 11.619 | 50 | 71 | 85.57 |
| Ebola virus | Virus | 0.09 | 8 | 100 | 100.00 |
| Echovirus | Virus | 0.024 | 20 | 100 | 99.89 |
| E. coli | Virus | 0.5 | 7 | 100 | 100.00 |
| Enterobacter cloacae | Bacteria | 1.414 | 24 | 100 | 100.00 |
| Enterococcus | Bacteria | 1.414 | 24 | 100 | 100.00 |
| Enterococcus faecalis | Bacteria | 0.707 | 11 | 100 | 100.00 |
| Francisella tularensis | Bacteria | 0.2 | 4 | 91 | 91.49 |
| Fusarium | Fungi | 11.225 | 50 | 92 | 96.23 |
| Haemophilus influenzae | Bacteria | 0.285 | 4 | 100 | 100.00 |
| Haemophilus parainfluenzae | Bacteria | 1.732 | 30 | 100 | 99.99 |
| Hantaan virus | Virus | 0.096 | 7 | 100 | 100.00 |
| Helicobacter pylori | Bacteria | 2.1 | 35 | 100 | 100.00 |
| Histoplasma capsulatum | Fungi | 2.236 | 36 | 99 | 99.56 |
| Influenza A virus | Virus | 0.098 | 7 | 100 | 100.00 |
| Junin virus | Virus | 0.122 | 6 | 100 | 100.00 |
| Klebsiella pneumoniae | Bacteria | 0.671 | 10 | 100 | 100.00 |
| Lassa virus | Virus | 0.122 | 6 | 100 | 100.00 |
| LCV | Virus | 0.087 | 8 | 100 | 100.00 |
| Legionella pneumophila | Bacteria | 0.52 | 7 | 100 | 100.00 |
| Listeria monocytogenes | Bacteria | 0.707 | 11 | 99 | 98.98 |

Table 4: Combined UV + Filter Removal Rates

| Microbe | Type | Size μm | Filter % | UV Rate % | Total % |
|----------------------------|----------|--------------------|----------|-----------|---------|
| Marburg virus | Virus | 0.039 | 15 | 100 | 100.00 |
| Measles virus | Virus | 0.158 | 5 | 100 | 100.00 |
| MERS virus | Virus | 0.11 | 6 | 89 | 90 |
| Mucor | Fungi | 7.071 | 50 | 95 | 98 |
| Mumps virus | Virus | 0.164 | 5 | 100 | 100 |
| Mycobacterium avium | Bacteria | 1.118 | 19 | 100 | 100 |
| Mycobacterium kansasii | Bacteria | 1.118 | 19 | 100 | 100 |
| Mycobacterium tuberculosis | Bacteria | 0.637 | 9 | 100 | 100 |
| Mycoplasma pneumoniae | Bacteria | 0.177 | 5 | 100 | 100 |
| Neisseria meningitidis | Bacteria | 0.775 | 12 | 100 | 100 |
| Nocardia asteroides | Bacteria | 1.118 | 19 | 100 | 100 |
| Norwalk virus | Virus | 0.029 | 18 | 97 | 98 |
| Parainfluenza virus | Virus | 0.194 | 4 | 100 | 100 |
| Panovirus B19 | Virus | 0.022 | 21 | 100 | 100 |
| Penicillium | Fungi | 3.262 | 44 | 60 | 78 |
| Proteus mirabilis | Bacteria | 0.494 | 7 | 100 | 100 |
| Pseudomonas aeruginosa | Bacteria | 0.494 | 7 | 100 | 100 |
| Reovirus | Virus | 0.075 | 9 | 99 | 99 |
| RSV | Virus | 0.19 | 5 | 100 | 100 |
| Rhinovirus | Virus | 0.023 | 21 | 99 | 99 |
| Rhizopus | Fungi | 6.928 | 50 | 93 | 96 |
| Rickettsia prowazeki | Bacteria | 0.6 | 9 | 100 | 100 |
| Rotavirus | Virus | 0.073 | 9 | 100 | 100 |
| Rubella virus | Virus | 0.061 | 11 | 67 | 71 |
| Salmonella typhi | Bacteria | 0.806 | 13 | 100 | 100 |
| SARS virus | Virus | 0.11 | 6 | 100 | 100 |
| Serratia marcescens | Bacteria | 0.632 | 9 | 100 | 100 |
| Stachybotrys chartarum | Fungi | 5.623 | 49 | 12 | 55 |
| Staphylococcus aureus | Bacteria | 0.866 | 14 | 100 | 100 |
| Staphylococcus epidermidis | Bacteria | 0.866 | 14 | 100 | 100 |
| Streptococcus pneumoniae | Bacteria | 0.707 | 11 | 77 | 80 |
| Streptococcus pyogenes | Bacteria | 0.894 | 14 | 100 | 100 |
| Trichophyton | Fungi | 4.899 | 49 | 71 | 85 |
| Ustilago | Fungi | 5.916 | 50 | 46 | 73 |
| VZV | Virus | 0.173 | 5 | 100 | 100 |
| Yersinia pestis | Virus | 0.707 | 11 | 100 | 100 |



Images during lab testing

PROOF OF EFFECTIVENESS

Tests conclusively support that UV Angel Air treats bacteria, fungus and viruses in the air including: **Gram negative and gram-positive bacteria, fungal pathogens and viral surrogates.**

The UV Angel Air results showed laboratory elimination rates up to **99.99%**.

Sources: Centers for Disease Control and Prevention. 2007. Guideline for Isolation Precautions: Preventing Transmission of Infectious Agents in Healthcare Settings. Available from: <http://www.cdc.gov/hicpac/2007IP/2007isolationPrecautions.html>. Accessed 26 August 2016 2. Tina Ethington, MSN, RN, CEN, NE-BC, Sherry Newsome, BSN, RN, MBA/MNA, Jerri Waugh, BSN, RN, MBA/MHA, Linda D. Lee, DrPH, MBA, Cleaning the air with ultraviolet germicidal irradiation lessened contact infections in a long-term acute care hospital, American Journal of Infection Control, December 2017 3. Douglas W. Kane, MD, Cynthia Finley RRT, Diane Brown RRT, Linda Lee PhD, UV-C Light and Infection Rate in a Long Term Care Ventilator Unit, May 23, 2016 4. Don Guimera, MSN, RN, CIC, CCRP, FAPIC, Jean Trzil, PharmD, Joy Joyner, RN, CIC, Nicholas D. Hysmith, MD, FAAP, Effectiveness of a shielded UV-C air disinfection system in an inpatient pharmacy of a tertiary care children's hospital, American Journal of Infection Control, August 2017 5. Linda D. Lee, DrPH, MBA, Surface and air: What impact does UV-C at the room level have on airborne and surface bacteria? Canadian Journal of Infection Control, Summer 2017 6. Lee. Report on the Performance of the UV Angel Air *Walker CM, Ko G. Effect of ultraviolet germicidal irradiation on viral aerosols. Environ. Sci. Technol. 2007, 41, 15, 5460-5465 Weiss M, Horzinek MC. Resistance of Berne virus to physical and chemical treatment. Vet Microbiol. 1986;11(1):21-41-49. doi:10.1016/0378-1135(86)90005-2 Hirano N, Hino S, Fujiwara K. Physico-chemical properties of mouse hepatitis virus (MHV-2) grown on DBT cell culture. Microbiol Immunol. 1978;22(7):377-90. Sakinimit M1, Inatsuki I, Sugiyama Y, Yagami K. Virucidal efficacy of physico-chemical treatments against coronaviruses and parvoviruses of laboratory animals. Jikken Dobutsu. 1998 Jul;37(3):341-5. Duan SM, et al. Stability of SARS coronavirus in human specimens and environment and its sensitivity to heating and UV irradiation. Biomed Environ Sci. 2003 Sep;16(3):246-55. Darnell ME, et al. Inactivation of the coronavirus that induces severe acute respiratory syndrome, SARS-CoV. J Virol Methods. 2004 Oct;121(1):85-91. *Kariwa H1, Fujii N, Takahima J. Inactivation of SARS coronavirus by means of povidone-iodine, physical conditions, and chemical reagents. Jpn J Vet Res. 2004 Nov;52(3):105-12.